Great nutraceutical potential of bioactive compounds from *Beta vulgaris cicla* and *rubra*

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Beta vulgaris subsp. *cicla* (BVc, leaf beet) and *Beta vulgaris* var. *rubra* (BVr, red beetroot) belong to the *Amaranthaceae* family and have been used for centuries as food and medicinal plants. The main bioactive phytochemicals of BVr are the betalains, a group of water-soluble pigments derived from betalamic acid, which are divided into two classes: the yellow/orange-coloured betaxanthins (BX) and the red/ violet-coloured betacyanins (BC). The seeds, leaves and roots of BVc are rich in phenolic acids and apigenin-derived flavonoids, namely vitexin, vitexin-2-O-rhamnoside (VOR) and vitexina-2-O-xyloside (XVX). We isolated BVc and BVr phytochemicals in our laboratory and tested them individually and in combination for their anticancer and anti-inflammatory activity. In cancer cells, vitexin flavonoids were able to induce intrinsic apoptosis, while betalains induced extrinsic apoptosis. Combinations of two or three molecules exhibited synergistic antioxidant, anti-inflammatory and anticancer activity, particularly towards hepatic, intestinal and urinary bladder tumours.

Keywords Anticancer Anti-inflammatory Antioxidants Betalains Nutraceutical products Vitexin flavonoids

Introduction

Beta vulgaris subsp. *vulgaris* is a herbaceous biennial plant belonging to the order of the *Caryophyllales*, in the family of the *Amaranthaceae*, and in the *Betoideae* subfamily [1]. *Beta vulgaris* has a diploid karyotype with nine pairs of chromosomes [2].

The flowers are produced in large inflorescences in the second year, at the apex of the stem. The fruit is an aggregate of two or more fruits, forming an irregular body which produces irregular brown seeds, with two cotyledons and one embryo (Fig. 1) [1].

The leafy beet group was domesticated first in the Mediterranean region and then introduced into the Middle East, India and China. Beet is now cultivated worldwide in temperate regions, with leafy beet preferring warmer temperatures than beetroot [3].

Taxonomically, *Beta vulgaris* is divided into four important crops (Table 1). *Beta vulgaris* subsp. *cicla* (BVc, leaf beet),

¹Department of Biomolecular Sciences, University of Urbino Carlo Bo, Via Saffi 2, 61029 Urbino (PU), Italy also called chard or spinach beet and grown for its leaves, is an important economic crop in many regions of Italy. *Beta vulgaris* var. *rubra* (BVr, red beetroot) is widely cultivated in Northern and Central Italy for its dark red, yellow or white roots. *Beta vulgaris* var. *altissima* (sugar beet) was widely grown in Italy until the 1980s for table sugar production but was then abandoned as unprofitable, as was fodder beet.

Cultivated and wild-type *Beta vulgaris* species have long been used in folk medicine. Leaf beet and beetroot are used to stimulate the immune system, protect bone, and combat inflammation, liver intoxication and gastrointestinal disease [1].

The juice and leaves of red beetroot are used in traditional medicine for genito-urinary and intestinal tumours, and to protect the gastro-intestinal tract [4]. The naturopath Rudolf Breuss developed a liquid formulation, with depurative and chemopreventive properties, based on red beetroot and other vegetables.

In this review, we describe the phenolic nutrients with pharmacological importance in *Beta vulgaris* subsp. *cicla* (BVc, leaf beet) and *Beta vulgaris* var. *rubra* (BVr, red beet-root) to stress their importance as bioactive phytochemicals for use in nutraceutical products.

Group	Taxon	Common name	Uses	Italian name
Leaf beet	Beta vulgaris subsp. vulgaris, convar. cicla	Swiss chard	Leaves and stalk boiled or steamed Medicinal plant	Bieta a coste
	<i>Beta vulgaris</i> subsp. <i>vulgaris,</i> convar. <i>cicla,</i> var. <i>cicla</i>	Spinach beet	Whole leaves boiled as spinach or raw in salads Medicinal plant	Bieta erbetta da taglio
Fodder beet	<i>Beta vulgaris</i> subsp. <i>vulgaris,</i> convar. <i>crassa</i>	Mangelwurzel	Raw as animal food	Bietola da radici
Sugar beet	<i>Beta vulgaris</i> subsp. <i>vulgaris,</i> convar. <i>vulgaris,</i> var. altissima	Sugar beet	Commercial crop for industrial sugar production	Barbabietola da zucchero
Garden beet	<i>Beta vulgaris</i> subsp. <i>vulgaris,</i> convar. <i>vulgaris,</i> var. <i>vulgaris</i>	Red beetroot, garden beet	Boiled or steamed as a food, centrifuged as a juice Medicinal plant	Bietola rossa

 Table 1 - Taxonomy of beet cultivars with their uses

Nutritional aspects

BVc leaves and BVr roots are usually eaten boiled or steamed. BVc is rich in minerals, such as Fe, K, Ca, P and Mg, and numerous vitamins such as B3, B5, B9, A, C and folic acid. Phenolic acids, flavonoids and carotenoids, fibre and saponins are present as well [3]. BVr is also consumed as a crude juice containing Ca, P, Fe, B vitamins and polyphenols [5]. Steaming allows the retention of most vitamins and polyphenols, while boiling releases most of the nutrients into the cooking water [6–8].

Patients with bladder problems must use caution when consuming BVc and BVr because of the presence of up to 1% oxalates in second-year plants. Variable amounts of geosmin, a bicyclic alcohol, are also present in BVr [9]. Geosmin has an unpleasant earthy flavour, which is mostly lost when the vegetable is boiled in plenty of water [5].

Pharmacology

BVc and BVr contain many phytochemicals providing pharmacological effects. Earlier studies by Ozsoy-Sacan *et al* [10] and Sener *et al* [11] demonstrated that BVc leaf extract exhibited hypoglycaemic properties in diabetic animals. Research to discover the components responsible for this pharmacological effect tentatively identified saponins, which inhibit gluconeogenesis and glycogenolysis [12]. However, the definitive mechanism responsible for the hypoglycaemic effect remains unknown and further investigation is required. In recent years, the wide use of separation technology, in particular automatic medium pressure liquid chromatography (MPLC) and cheap new stationary phases has allowed many laboratories to prepare milligram amounts of individual phytochemicals in order to study their specific biological properties. Using a dereplication process, we extracted the secondary metabolites using ethanol and then isolated the individual phytochemicals from BVc and BVr using liquid–liquid fractionation and MPLC. We identified their chemical formulas using HPLC-MS and NMR and tested their biological activities in tumour cell lines. We used the following cancer cell lines: breast, MDA-MB-231, MCF-7; colon, LoVo, CaCo-2, RKO; hepatic, HepG2, Hep3B; and urinary bladder, T24. In parallel, we performed similar experiments with normal cell lines, such as human fibroblasts (HF), human keratinocytes (NCTC-2544) and human lymphocytes, in order to determine if the action against cancer cells was replicated in normal cells [5].

The chemical structures of the active phytochemicals of BVc and BVr studied in our laboratory are given in Fig. 1.

The seeds, leaves and roots of leaf beet are rich in phenolic acids such as siringic, caffeic and vanillic, as well as flavonoids derived from apigenin (Fig. 1), such as vitexin, vitexin-2-O-rhamnoside (VOR) and vitexin-2-O-xyloside (VOX) [5], and also catechin, myricetin and quercetin [3].

The main molecules in BVr are betalains, a group of watersoluble phenolic pigments derived from betalamic acid. They are subdivided into two classes: the yellow/orange betaxanthins (BX) and the red/violet betacyanins (BC). The main betacyanin in beetroot is betanin with its aglycone betanidin, while the most abundant betaxanthin is vulgaxanthin I (Fig. 1) [13, 14].

The betalains were isolated in the 1930s and intensively studied in the 1980s and 1990s [13, 15–17]. These pigments have recently been receiving increased attention as they are quite stable at the pH values of many foods and are used as an additive dye in various food products, particularly cheese, tomato paste, sauces, ice cream, sweets, breakfast cereals and desserts.

The European Food Safety Authority (EFSA) allows the use of red beetroot dye, which has the e-number E162 [18]. This

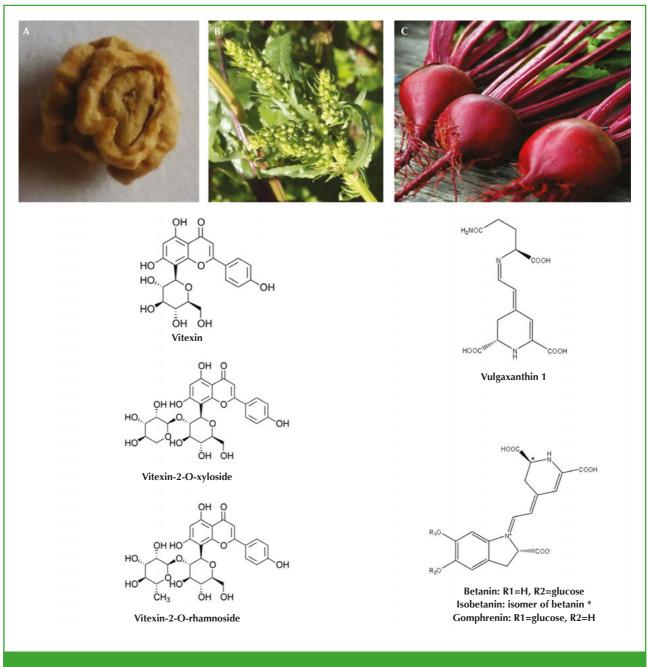


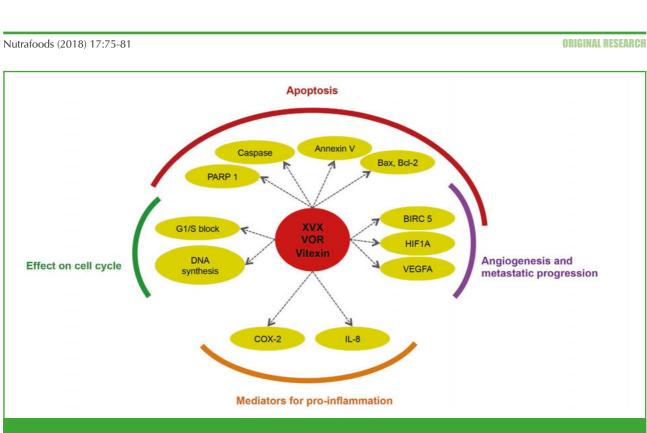
Figure 1 - Seed, flowers and roots of *Beta vulgaris* and structure of vitexin flavonoids and betalains: (**A**) seed of *Beta vulgaris* subsp. *cicla*; (**B**) flowers and fruits of *Beta vulgaris* var. *rubra*; (**C**) roots of *Beta vulgaris* var. *rubra*, cv. Detroit

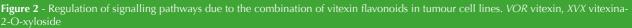
dye consists of up to 90% betalains, and therefore exerts a healthy effect due to the high antioxidant capacity of the pigment [14]. The amount of dye added to foods reaches 10–15 mg/100 g food, thus increasing antioxidant capacity and protecting the body during post-prandial oxidative stress [19].

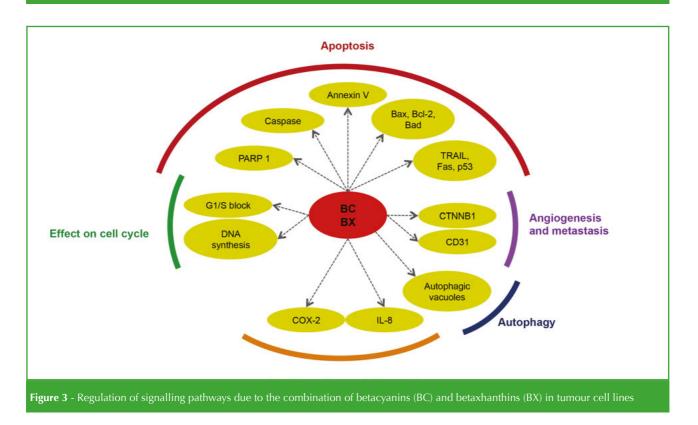
The pharmacological effects of the BVc and BVr bioactive molecules are summarized in Figs. 2 and 3.

Figure 2 shows the changes in key molecules involved in different signalling pathways due to vitexin flavonoids. The inhibition of DNA synthesis and cell cycle progression was detected by flow cytometry, showing a reduction in the number of cells in the S-phase due to arrest of the cell cycle in G1 [20]. Experiments with tritiated thymidine were also performed and results were expressed as % of DNA synthesis inhibition [21].

Induction of apoptosis was evaluated using (a) fluorescent detection of annexin V bound to apoptotic cells, and quantitation by flow cytometry [14, 22], (b) detection of an increase in pro-apoptotic proteins, such as Bax and a decrease in the anti-apoptotic Bcl family proteins [14, 22], (c) detection of an increase in cleaved PARP by measuring PARP1 fragments (PARP is the enzyme which contributes to DNA repair after oxidative stress and is cleaved during apopto-







sis into two fragments, one of which, called PARP1, is an excellent marker of apoptosis [14]), and (d) detection of an increase in the caspases, particularly caspase 3, which is the executioner caspase responsible for the cleavage of many key proteins. The enzymatic assay of caspase 3 measures the progression of the apoptotic pathways in tumour cells [14, 22].

As apoptosis can be blocked by an increase in the concentration of inhibitor of apoptotic proteins (IAP), downregulation of these pro-survival markers in tumour cells means that ultimately apoptosis is accelerated. We observed downregulation of the mRNA expression level of the BIRC5 gene which encodes an IAP protein called Survivin. Moreover, vitexin flavonoids were found to downregulate mRNA levels of HIF1 α and VEGF-A, which are regulators of angiogenesis and controllers of tumour cell proliferation and invasiveness [23]. We also observed depletion of the pro-inflammatory markers COX-2 and IL-8, which are important for triggering apoptosis. To summarize, vitexin flavonoids show multitargeted anticancer activity, which triggers apoptosis and downregulates proliferation, survival and angiogenesis factors in tumour cells [5].

Figure 3 shows the key molecules involved in the signalling pathways regulated by betalains. The inhibition of proliferation and the induction of apoptosis were detected using the same methods as reported above for vitexin flavonoids. In the presence of BC and BX, the apoptosis occurred through the extrinsic pathway and in addition there was a dramatic decrease in the mRNA expression levels of COX-2 and IL-8, which was greater than the decrease caused by the vitexin flavonoids [14]. Inhibition of angiogenesis has also been found by other authors [24] with depletion of the expression levels of cluster of differentiation 31, also called CD31, which is involved in angiogenesis and tumour progression. We also detected downregulation of the mRNA levels of CT-NNB1 which expresses β -catenin, a protein involved in prosurvival mechanisms [22].

Finally, additive pro-apoptotic signals due to betalains, which modified Bad, TRAIL, Fas and p53 expression levels, as well as the formation of autophagic vacuoles, were described by Nowacki *et al* [25].

In summary, BC and BX and their combination exert multiple anticancer effects by targeting several signalling pathways with resulting inhibition of tumour cell proliferation and inflammation.

Using the ORAC method, we determined that the chemical antioxidant capacity of vitexin flavonoids was approximately 20,000 µmolTE/g [5], while the ORAC values of BX and BC were approximately 10,000 µmolTE/g [14].

The values of the key markers involved in tumour cell progression indicate that the phytochemical combination XVX+BC+BX is the most efficacious followed by XVX+BC, due to the synergistic effects among the pathways described in Figs. 2 and 3.

Interestingly, our studies and those of other authors [5] showed that in cancer cell lines, vitexin flavonoids trigger the production of reactive oxygen species (ROS) and induce apoptosis through the intrinsic apoptosis pathway, which involves mitochondrial damage and cytochrome c release. The betalains behave as antioxidants within the cells, but trigger the extrinsic apoptosis pathway, which involves Fas and Fas ligand, with recruitment of Fas-associated proteins and acti-

vation of caspase 8, which in turn activates the executioner caspase 3 [5].

This means that a cancer prevention strategy using non-toxic phytochemicals (i.e., chemoprevention) could use a combination of vitexin flavonoids and betalains which activate both the intrinsic and extrinsic apoptotic pathways and block the salvage pathway of tumour cell lines.

Bioavailability

Bioavailability and bioaccessibility are both important for the study of the biological effects of phytochemicals. These topics are widely discussed in the literature and are the first factors to be considered when the results from in vitro experiments are translated to in vivo models.

In our laboratory, we evaluated the bioavailability of vitexin flavonoids using an ex vivo system, where a flavonoid was incubated with red blood cells in the presence of 2',7'-dichlorofluorescein diacetate (DCFH-DA), with subsequent measurement of cellular antioxidant activity [26]. This experiment showed that the flavonoid is able to enter red blood cells. A second method utilized polyclonal antibodies for the detection of vitexin flavonoids and their metabolites in the blood of Balb/c mice. Vitexin flavonoids were found in the plasma of mice fed 170 mg/kg of VOX [3].

Wang *et al* demonstrated that after oral administration of vitexin to experimental animals, the compound and its metabolites were found in the intestine and liver, as well as at very low concentrations in the brain and adipose tissue [27]. Similar results were found by Yan *et al* [28] and Liang *et al* [29]. These and other experiments [5], based on oral dose administration, concluded that the bioavailability of the vitexin flavonoid was approximately 3%.

Studies on the intra-caecal administration of XVX in a rat model demonstrated that it was quickly absorbed from the enterocytes and inserted into the entero-hepatic circulation [30]. Other authors evaluated the contribution of bacteria to the transformation of flavonoid C-glycosides by incubating these phytochemicals in the presence of isolated bacterial strains. The authors showed that the same bacteria are able to deglycosylate the C-glycosidic bond, making the molecular fragments bioavailable [31].

To summarize, vitexin is absorbed well by the intestine, likely by passive diffusion. The sugar moiety of XVX and VOR probably limits absorption, but bacterial metabolism and entero-hepatic recirculation prolong the time the molecules remain in the liver and intestines, possibly enhancing their pharmacological effects in these two compartments.

REFERENCES

Most phytochemicals have poor bioavailability due to their low capacity for absorption through the cell membrane or for extrusion from cells because of the multidrug resistance (MDR) system. Therefore, combination with other phytochemicals able to inhibit the MDR system should increase the bioavailability of the vitexin flavonoids.

Betalain bioavailability was studied in both animals and humans after the ingestion of beetroot juice [32, 33]. Betacyanins were found in the urine, mostly as metabolized products, but bioavailability was low [34, 35]. Tesoriere *et al* demonstrated in an in vitro model which mimics the gut functional barrier, that betanin and indicaxanthin were absorbed unmetabolized via paracellular transport [15].

To summarize, most of the experiments cited above sought to identify the few known metabolites of betalains and vitexin flavonoids in blood and organs. Only after complete elucidation of their metabolic pattern, through phase 1 and 2 reactions, will it be possible to draw definitive conclusions on the amount of molecules absorbed.

Conclusions

The combination of vitexin flavonoids and betalains has been shown to be very efficient for combating inflammation and tumour cell proliferation. The future development of functional foods and nutraceutical products with chemopreventive efficacy must be based on the selection of precise BVc and BVr cultivars and standardized automatic methods to obtain ethanolic extracts with high titres of vitexin flavonoids and betalains. Purification methods used in the laboratory to increase the percentage of the bioactive molecules should be scaled up and also adapted for use under food grade conditions. Toxicity tests and studies on animal models receiving long-term treatment should be carried out, and pharmacokinetic studies to detect metabolites in the different organs must also be conducted. These steps could result in an application being made to the EFSA for the listing of vitexin flavonoids and betalains as novel foods. Combinations of these molecules could be used to develop nutraceutical products to reduce the social and economic burden of cancer and other disorders associated with aging. The development of such products could lead to the production, industrial transformation and commercialization of beet, thus offering economic opportunities to the agri-food sector.

Conflict of interest

The authors declare that they have no conflict of interest.

- Duke JA (1983) Handbook of energy crops. http://www.hort.purdue.edu/newcrop/duke_energy/dukeindex.html (accessed 29 May 2018)
- Kong W, Yang S, Wang Y, Bendahmane M, Fu X (2017) Genomewide identification and characterization of aquaporin gene family in *Beta vulgaris*. PeerJ 5:e3747
- Ninfali P, Angelino D (2013) Nutritional and functional potential of Beta vulgaris cicla and rubra. Fitoterapia 89:188–199
- 4. Chiej R (1984) Encyclopaedia of medicinal plants. MacDonald, London
- Ninfali P, Antonini E, Frati A, Scarpa ES (2017) C-glycosyl flavonoids from *Beta vulgaris cicla* and betalains from *Beta vulgaris rubra*: antioxidant, anticancer and antiinflammatory activities–a review. Phytother Res 31:871–884
- Ninfali P, Bacchiocca M (2003) Polyphenols and antioxidant capacity of vegetables under fresh and frozen conditions. J Agric Food Chem 51:2222–2226
- Gil MI, Ferreres F, Tomas-Barberan FA (1999) Effect of postharvest storage and processing on the antioxidant constituents (flavonoids and vitamin C) of fresh-cut spinach. J Agric Food Chem 47:2213– 2217
- Ninfali P, Mea G, Giorgini S, Rocchi M, Bacchiocca M (2005) Antioxidant capacity of vegetables, spices and dressings relevant to nutrition. Br J Nutr 93:257–266
- Lu G, Edwards CG, Fellman JK, Mattinson DS, Navazio J (2003) Biosynthetic origin of geosmin in red beets (*Beta vulgaris* L.). J Agric Food Chem 51:1026–1029
- Ozsoy-Sacan O, Karabulut-Bulan O, Bolkent S, Yanardag R, Ozgey Y (2004) Effects of chard (*Beta vulgaris* L. var *cicla*) on the liver of the diabetic rats: a morphological and biochemical study. Biosci Biotechnol Biochem 68:1640–1648
- Sener G, Sacan O, Yanardag R, Yanoglu-Dulger G (2002) Effects of chard (*Beta vulgaris* L. var. *cicla*) extract on oxidative injury in the aorta and heart of streptozotocin-diabetic rats. J Med Food 5:37–42
- Massiot G, Dijoux MG, Lavaud C, Le Men-Olivier L, Connolly JD, Sheeley DM (1994) Seco-glycosides of oleanolic acid from *Beta vul*garis. Phytochemistry 37:1667–1670
- Stintzing FC, Carle R (2004) Functional properties of anthocyanins and betalains in plants, food, and in human nutrition. Trends Food Sci Technol 15:19–38
- Farabegoli F, Scarpa ES, Frati A, Serafini G, Papi A, Spisni E *et al* (2017) Betalains increase vitexin-2-O-xyloside cytotoxicity in CaCo-2 cancer cells. Food Chem 218:356–364
- Tesoriere L, Gentile C, Angileri F, Attanzio A, Tutone M, Allegra M et al (2013) Trans-epithelial transport of the betalain pigments indicaxanthin and betanin across Caco-2 cell monolayers and influence of food matrix. Eur J Nutr 52:1077–1087

- Stintzing FC, Herbach KM, Mosshammer MR, Carle R, Yi WG, Sellappan S *et al* (2005) Color, betalain pattern, and antioxidant properties of cactus pear (*Opuntia* spp.) clones. J Agric Food Chem 53:442–451
- 17. Stintzing FC, Carle R (2005) Cactus stems (*Opuntia* spp.): a review on their chemistry, technology, and uses. Mol Nutr Food Res 49:175–194
- EU (2015) Scientific Opinion on the re-evaluation of beetroot red (E 162) as a food additive. EFSA J 13:4318–4373
- Prior RL, Xianli W (2013) Diet antioxidant capacity: relationships to oxidative stress and health. Am J Biomed Sci 5:126–139
- 20. Gennari L, Felletti M, Blasa M, Angelino D, Celeghini C, Corallini A *et al* (2011) Total extract of *Beta vulgaris* var. *cicla* seeds versus its purified phenolic components: antioxidant activities and antiproliferative effects against colon cancer cells. Phytochem Anal 22:272–279
- 21. Ninfali P, Bacchiocca M, Antonelli A, Biagiotti E, Di Gioacchino AM, Piccoli G *et al* (2007) Characterization and biological activity of the main flavonoids from Swiss chard (*Beta vulgaris* subspecies *cycla*). Phytomedicine 14:216–221
- 22. Scarpa ES, Emanuelli M, Frati A, Pozzi V, Antonini E, Diamantini G *et al* (2016) Betacyanins enhance vitexin-2-O-xyloside mediated inhibition of proliferation of T24 bladder cancer cells. Food Funct 7:4772–4780
- Baba Y, Nosho K, Shima K, Irahara N, Chan AT, Meyerhardt JA *et al* (2010) HIF1A overexpression is associated with poor prognosis in a cohort of 731 colorectal cancers. Am J Pathol 176:2292–2301
- 24. Zhang Q, Pan J, Wang Y, Lubet R, You M (2013) Beetroot red (betanin) inhibits vinyl carbamate- and benzo(a)pyrene-induced lung tumorigenesis through apoptosis. Mol Carcinog 52:686–691
- 25. Nowacki L, Vigneron P, Rotellini L, Cazzola H, Merlier F, Prost E et al (2015) Betanin-enriched red beetroot (*Beta vulgaris* L.) extract induces apoptosis and autophagic cell death in MCF-7 cells. Phyto-

ther Res 29:1964-1973

- 26. Blasa M, Angelino D, Gennari L, Ninfali P (2011) The cellular antioxidant activity in red blood cells (CAA-RBC): a new approach to bioavailability and synergy of phytochemicals and botanical extracts. Food Chem 125:685–691
- Wang YJ, Han CH, Leng AJ, Zhang WJ, Xue HF, Chen YH *et al* (2012) Pharmacokinetics of vitexin in rats after intravenous and oral administration. Afr J Pharm Pharmacol 6:2368–2373
- Yan C, Liu H, Lin L (2013) Simultaneous determination of vitexin and isovitexin in rat plasma after oral administration of *Santalum album* L. leaves extract by liquid chromatography tandem mass spectrometry. Biomed Chromatogr 27:228–232
- 29. Liang M, Xu W, Zhang W, Zhang C, Liu R, Shen Y *et al* (2007) Quantitative LC/MS/MS method and in vivo pharmacokinetic studies of vitexin rhamnoside, a bioactive constituent on cardiovascular system from hawthorn. Biomed Chromatogr 21:422–429
- Angelino D, Berhow M, Ninfali P, Jeffery EH (2013) Caecal absorption of vitexin-2-O-xyloside and its aglycone apigenin, in the rat. Food Funct 4:1339–1345
- Kim M, Lee J, Han J (2015) Deglycosylation of isoflavone C-glycosides by newly isolated human intestinal bacteria. J Sci Food Agric 95:1925–1931
- Clifford T, Howatson G, West DJ, Stevenson EJ (2015) The potential benefits of red beetroot supplementation in health and disease. Nutrients 7:2801–2822
- Clifford T, Constantinou CM, Keane KM, West DJ, Howatson G, Stevenson EJ (2017) The plasma bioavailability of nitrate and betanin from *Beta vulgaris rubra* in humans. Eur J Nutr 56:1245–1254
- 34. Kanner J, Harel S, Granit R (2001) Betalains--a new class of dietary cationized antioxidants. J Agric Food Chem 49:5178–5185
- 35. Frank T, Stintzing FC, Carle R, Bitsch I, Quaas D, Strass G et al (2005) Urinary pharmacokinetics of betalains following consumption of red beet juice in healthy humans. Pharmacol Res 52:290–297